10X Tris-BES Running Buffer

1. Catalog No. KTQ030

2. Quantity 500 ml

3. Storage & Stability Store at R.T

4. Description 10X Tris-BES Running Buffer is used for running Protein samples for SDS PAGE analysis on Polyacrylamide gels.

5. Recommended Dilution Add 1 volume of 10X Tris-BES Running Buffer to 9 volumes of D.W..

6. Protocol

1. Prepare your sample by adding one part of the appropriate Tris-BES Sample Buffer (2X), to one part of sample and mix well. For denaturing conditions, heat the sample at 95°C for 5 minutes. For reducing conditions, add 0.2 ml of DTT Reducing reagent or 0.05 ml of 2-Mercaptoethanol to 1 ml of Tris-BES Sample Buffer (2X) before use.

2. Dilute the Tris-BES Running Buffer (10X), 1:9 with deionized water before use. Fill the upper and lower buffer chambers of the Ezcell with appropriate amounts of running buffer. For reducing conditions, just prior to the run, prepare an upper buffer (cathode) by adding 0.5 ml of Antioxidant (400X) to 200 ml of Tris-BES Running Buffer (1X).

3. Run the gel according to the following running conditions.
   Note: Runtime is dependent on the gel percentage. The run is complete when the Coomassie blue tracking dye reaches the bottom of the gel.

Current	60 mA constant	
Approx. Voltage	Start End	160-220 V / 1.0mm gel 250-290 V / 1.0mm gel
Approx. Run Time	37 minutes	

4. After the run, remove the gel from the cassette.

5. Fix, stain or transfer as desired.