10X Tris-Glycine Native Running Buffer

1. Catalog No. KTG030N

2. Quantity 500 ml

3. Storage & Stability Store at R.T.

4. Description 10X Tris-Glycine Running Native Buffer is used for loading Protein samples for Native PAGE analysis on Polyacrylamide gels.

5. Recommended Add 1/10 volume of 10X Tris-Glycine Native Running Buffer to your protein sample tubes.

6. Loading

1. Tris-Glycine Native Sample Buffer does not contain SDS. Add one part of sample to one part of Tris-Glycine Native Sample Buffer (2X) and mix well. (Do not heat.)
2. Prepare 800ml 1X Tris-Glycine Native Running Buffer by adding 80ml of Tris-Glycine Native Running Buffer (10X) to 720ml of deionized water before use. Fill the upper and lower buffer chambers of EzCell with the appropriate amounts of running buffer. Make sure that the upper running buffer covers the sample wells completely.
3. Load sample into the wells. Typically 0.1-0.5ug of protein per band with Coomassie Blue staining will give optical band intensity.
4. Run the gel according to the following running conditions. Commonly a voltage of 125V constant is applied for Native PAGE gels, we recommend a voltage of 150V constant for this gel with Tris-Glycine Buffer.

Voltage	125V constant	
Approx. Current	Start	6-12 mA / 1.0mm gel
	End	3-6 mA / 1.0mm gel
Approx. Run Time	1-12 hour	

Turn off the power when the BPB dye is migrated to the end of the gel.

5. After the run, remove the gel from the cassette.
6. Fix, stain or transfer as desired.

*** Buffer composition**

Tris-Glycine Running Buffer (10x), 1L	
Tris base	0.25 M
Glycine	1.92 M