2X Tris-Glycine Native Sample Buffer

1. Catalog No. KTG020N

2. Quantity 20 ml

3. Storage & Stability Store at 4°C

4. Description 2X Tris-Glycine Native Sample Buffer is used for loading Protein samples for Native PAGE analysis on Polyacrylamide gels.

5. Recommended Loading Add 1/2 volume of 2X Tris-Glycine Native Sample Buffer to your protein sample tubes.

6. Protocol

1. **Tris-Glycine Native Sample Buffer does not contain reducing agent. For reducing conditions, add 0.25ml of 2-Mercaptoethanol or 0.1g of DTT to 5ml of Tris-Glycine Native Sample Buffer (2x) before use**.
2. Add one part of sample to one part of Tris-Glycine Native Sample Buffer (2x) and mix well. (Do not heat).
3. Prepare 800ml of Tris-Glycine Native Running Buffer (1x) by adding 80ml of Tris-Glycine Native Running Buffer (10x) to 720ml of deionized water before use. Fill the upper and lower buffer chambers of mini cell with the appropriate amounts of running buffer. Make sure that the upper running buffer covers the sample wells completely.
4. Load sample into the wells. Typically 0.1-0.5ug of protein per band with Coomassie Blue staining will give optical band intensity.
5. Run the gel according to the following running conditions. Commonly a voltage of 125V constant is applied for Native-PAGE (Laemmli) gels, we recommend a voltage of 125V constant for this gel with Tris-Glycine Native Buffer.

Voltage	125V constant	
Approx. Current	Start	6-12mA / 1.0mm gel
	End	3-6mA / 1.0mm gel
Approx. Run Time	1-12 hour	

Turn off the power when the BPB dye is migrated to the end of the gel.

6. After the run, remove the gel from the cassette.
7. Fix, stain or transfer as desired.

*** Buffer composition**

Tris-Glycine SDS Sample Buffer (1x)	
Tris-HCl, pH 6.8	63 mM
Glycerol	10 %
Bromophenol Blue	0.0025 %