

2X Aspartate Sample Buffer

- 1. Catalog No. KAS020
- 2. Quantity 20 ml

Loading

6. Protocol

- 3. Storage & Stability Store at 4°C
- **4. Description** 2X Aspartate Sample Buffer is used for loading Protein samples for SDS PAGE analysis on Polyacrylamide gels.
- **5. Recommended** Add 1 volume of 2X Aspartate Sample Buffer to your protein sample tubes.
 - 1. Aspartate Sample Buffer does not contain reducing agent. For reducing conditions, add 0.25ml of 2-Mercaptoethanol or 0.1g of DTT to 5ml of Aspartate Sample Buffer (2X) before use.
 - 2. Add one part of sample to one part of Aspartate Sample Buffer (2X) and mix well. Heat the sample at 85 $^\circ\!C$ for 2 minutes.
 - 3. Prepare 800ml 1X Aspartate Running Buffer by adding 80ml of Aspartate Running Buffer (10X) to 720ml of deionized water before use. Fill the upper and lower buffer chambers of EzCell with the appropriate amounts of running buffer. Make sure that the upper running buffer covers the sample wells completely.
 - 4. Load sample into the wells. Typically 0.1-0.5ug of protein per band with Coomassie Blue staining will give optical band intensity.
 - 5. Run the gel according to the following running conditions.

Voltage		100V constant
Approx. Current	Start	100mA / 1.0mm gel
	End	60mA / 1.0mm gel
Approx. Run Time		60 minutes

Turn off the power when the CBB dye is migrated to the end of the gel.

- 6. After the run, remove the gel from the cassette.
- 7. Fix, stain or transfer as desired.

* Buffer composition

Aspartate Sample Buffer (1x)		
Tris-HCI, pH 8.45	0.15 M	
Glycerol	12 %	
SDS	2 %	
Coomassie Blue G	0.003 %	