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## Notice

Check your package **insert & kit box for lot** specific instructions

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# KOMA ELISA Kit

## User Manual



LABISKOMA



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## Human TGF-beta 2 ELISA kit, pink-ONE

<b>Catalog No.</b>	<b>K0332120P</b>
<b>Lot No.</b>	<b>33232</b>
<b>Quantity</b>	<b>96 tests</b>
<b>Storage</b>	<b>4°C</b>
<b>Standard Range</b>	<b>31.25-2000 pg/ml</b>

### **[Important Notice]**

- **Please read this User Manual carefully prior to performing the assay.**
- **The reagent preparation method might be different from lot to lot, so please check the lot and follow the instructions given in this manual.**
- **Do not mix or interchange reagents between different lots.**
- **The kit is intended for Research Use Only.**

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## DESCRIPTION

This kit contains all the necessary reagents required for performing quantitative measurement of the protein from samples including serum, plasma, culture medium or other biological fluids in a sandwich ELISA format.

## KIT COMPONENTS

Component	Amount
Pre-Coated 96 well ELISA microplate	1 Plate
Biotinylated Affinity Purified Detection Antibody (Lyophilized)	2 EA
Recombinant Standard Protein (Lyophilized)	2 EA
Streptavidin-HRP Conjugate (0.6 ml)	1 EA
Assay Diluent (50 ml) : 1% BSA in PBS	1 EA
Assay Dileunt G (10 ml) : N/A	N/A
TMB or pink-ONE Solution (10 ml)	1 EA
Stop Solution (10 ml)	1 EA
Wash Buffer Concentrate (20X, 50 ml) to make 1 liter	1 EA
Plate Sealer	3 EA

## STORAGE AND STABILITY

- Store kit at 4°C immediately upon receipt.
- The shelf life of the kit is one year from date of shipment.
- Expiry of the kit is stated on labels.

## STANDARD RANGE

Standard Range                      31.25-2000 pg/ml

## SAMPLE PREPARATION

- Store all samples on ice after preparation and use immediately or aliquot and store at -80°C.
- Avoid repeated freeze-thaw cycles.

### 1) Cell culture supernatants

Centrifuge cell culture media at 1,500 rpm for 10 minutes at 4°C to remove particulates.

Immediately aliquot supernatants and store at -80°C.

### 2) Serum

Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 3,000 rpm for 10 minutes at 4°C to remove clots. Immediately aliquot supernatants and store at -80°C.

### 3) Plasma

Collect plasma using anti-coagulant (citrate, EDTA or heparin). Centrifuge samples at 3,000 rpm for 15 minutes at 4°C. Immediately aliquot supernatants and store at -80°C.

### 4) Sample activation and dilution

latent TGF-beta contained within samples are required activation.

Sample Activation Procedures	
Culture Supernatant	Serum/Plasma
1. Add 20 ul of 1 N HCl to 100 ul of Culture Supernatant.	1. Add 20 ul of 1 N HCl to 40 ul of Serum/Plasma.
2. Mix well by vortexing	2. Mix well by vortexing
3. Incubate for 10 min at room temperature.	3. Incubate for 10 min at room temperature.
4. The acidified sample neutralize by adding 20 ul of 1.2 N NaOH/0.5 M HEPES.	4. The acidified sample neutralize by adding 20 ul of 1.2 N NaOH/0.5 M HEPES.
5. Mix well by vortexing	5. Mix well by vortexing
6. Using this activated sample to assay immediately.	6. Activated sample dilute 20-fold with Assay diluent. (10 ul of activated sample + 190 ul of assay diluent)
7. The reading sample concentration must be multiplied by the dilution factor, 1.4.	7. Using this dilute-activated sample to assay immediately.

8. The reading sample concentration must be multiplied by the dilution factor, 40

Note : The kit Standards contain active recombinant TGF-beta. **do not activate the kit standards.**

**Use polypropylene test tubes.**

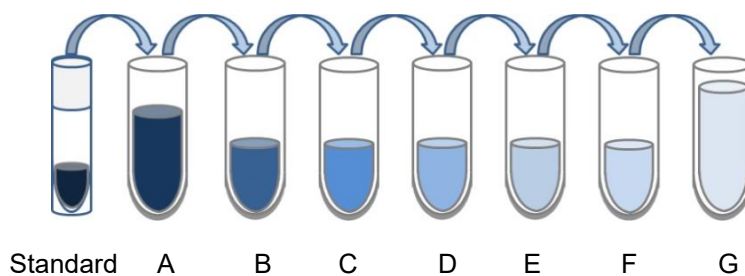
**Note:** Dilute the samples, based on the expected concentration of the analyte, to fall within the concentration range of the standards.

## REAGENT PREPARATION

- **Do not mix or substitute Assay Diluent from other kit lots.**
- **All reagents should be prepared right before use, and diluted solution should be used immediately.**

### 1) Standard Protein

Reconstitute 1 vial of Standard protein in 0.055 ml sterile water to a concentration of 25,000 pg/ml. Then dilute in Assay Diluent at 1:2 serial dilutions as follows. The standard diluent buffer serves as zero standard.



**Table 1.**

Step	Dilution Method		Concentration
Step A	0.04 ml of Standard	+ 0.46 ml of Assay Diluent	2000 pg/ml
Step B	0.25 ml of Step A	+ 0.25 ml of Assay Diluent	1000 pg/ml
Step C	0.25 ml of Step B	+ 0.25 ml of Assay Diluent	500 pg/ml
Step D	0.25 ml of Step C	+ 0.25 ml of Assay Diluent	250 pg/ml
Step E	0.25 ml of Step D	+ 0.25 ml of Assay Diluent	125 pg/ml

Step F	0.25 ml of Step E	+ 0.25 ml of Assay Diluent	62.5 pg/ml
Step G	0.25 ml of Step F	+ 0.25 ml of Assay Diluent	31.25 pg/ml

## 2) Detection Antibody

Reconstitute 1 vial of Detection Antibody in 0.275 ml sterile water, and dilute 1:40 in Assay Diluent.

**NOTE:** Reconstituted solutions are stable at -20°C for up to 1 month. Do not repeat freezing and thawing.

## 3) Streptavidin-HRP

Dilute the Streptavidin-HRP conjugate 1:20 in Assay Diluent.

## 4) Wash Buffer

Dilute the 20X Wash Buffer Concentrate in 1 L distilled water.

## ELISA PROCEDURE

1) **Washing:** Add 200 ul of Washing Solution to each well. Aspirate the wells to remove liquid and wash the plate 3 times using 300 ul of Washing Solution per well. After the last wash, invert plate to remove residual solution and blot on paper towel.

**NOTE:** Do not let the well dry completely and go immediately to the next step.

2) **Reaction:** Add 100 ul of standard, blank and sample to each well in duplicate. Cover the plate with the Plate Sealer. Incubate at room temperature for at least 2 hours.

3) **Washing:** Aspirate the wells to remove liquid and wash the plate 4 times as in step 1.

**NOTE:** Vigorous washing of the plate after incubation steps is essential to obtaining low background values.

4) **Detection:** Add 100 ul of the diluted detection antibody per well. Then cover the plate with the Plate Sealer. Incubate at room temperature for 2 hours.

5) **Washing:** Aspirate and wash plate 4 times as in step 1.

6) **Conjugates:** Add 100 ul of the diluted Streptavidin-HRP per well. Cover the plate with the Plate Sealer. Incubate 30 minutes at room temperature (or at 37°C for 30 minutes).

7) **Washing:** Aspirate and wash plate 4 times as in step 1.

8) **Color Development:** Add 100 ul of TMB or pink-ONE TMB solution to each well. Incubate at room temperature for a proper



color development. Add 100 ul of the stop solution to each well.

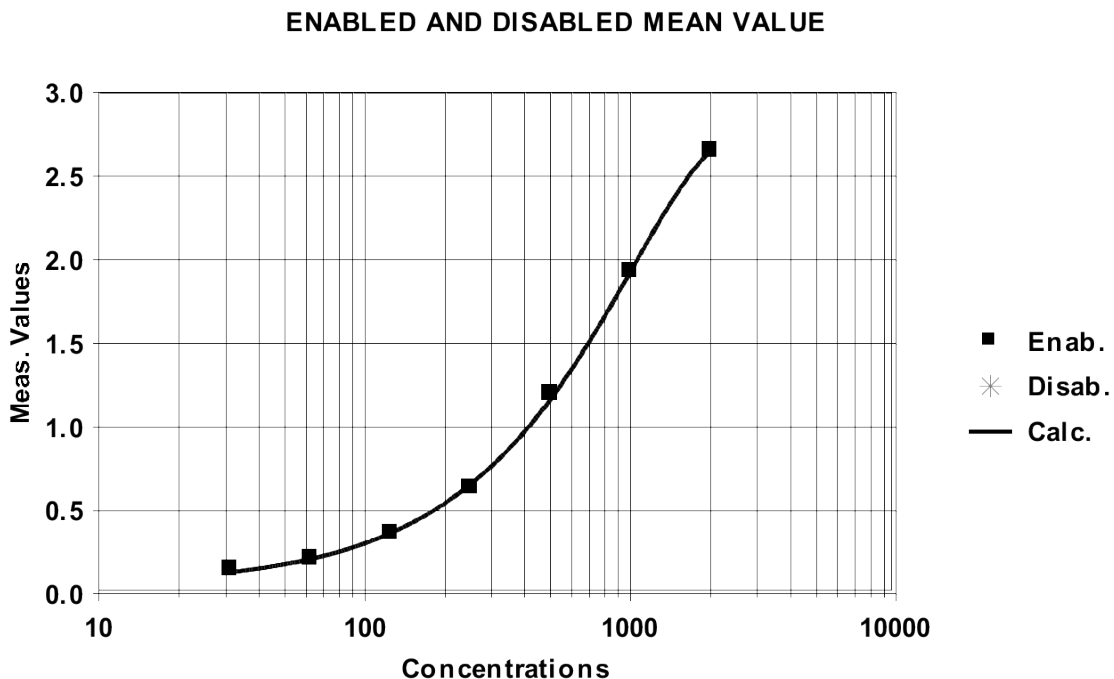
**NOTE:** Please be aware that the color may develop more quickly or more slowly than the recommended incubation time depending on the localized room temperature. Please monitor the color development to optimize the incubation time.

**NOTE:** Stop Solution ( $\text{H}_2\text{SO}_4$ ) is a caustic material. Eye, hand, face and clothing protection should be worn when handling this reagent.

9) **Reading:** Using a microplate reader, measure absorbance at 450 nm.

## CALCULATION OF RESULTS

Create a standard curve by reducing the data using ELISA reader's computer software capable of generating Standard curve-fit. A standard curve should be generated for each set of samples.



**Human TGF-beta 2 (pg/ml)**  
(3 minutes color development)

## CROSS REACTIVITY

To define the specificity of this ELISA, several proteins were tested for cross reactivity at 50 ng/ml.

Human : TGF-a, TGF-B1, TGF-B sRII, TGF-B3, TGF-B sRII/Fc Chimera,

Latent TGF-B1

Mouse : N/A

Rat : N/A

Others : Pig TGF-B1, Amphibian TGF-B5

## TROUBLESHOOTING

Problem	Probable Cause	Solution
<b>Low OD No Signal</b>	Reagents not fresh or contaminated	Ensure proper preparation of reagents.
	Incubation time not long enough	Ensure sufficient incubation times.
	Incubation temperature too low	Reagent solutions should be at RT before use.
	Stop solution not added	Addition of stop solution
<b>High OD</b>	Inadequate standard dilution	Ensure proper dilution of Standard.
	Inadequate incubation time of detection antibody, Streptavidin-HRP or Substrate	Decrease incubation time.
<b>Poor consistency of replicates</b>	Inadequate washing	Increase the stringency of washes.
	Inaccurate pipetting	Ensure accurate pipetting of volume and avoid air bubbles.
	Inadequate mixing of samples	Mix samples thoroughly before pipetting
	High particulate matter of samples	Mix samples thoroughly and remove particulates by centrifugation.
	Cross-well contamination	Use fresh plate sealers or pipette tips
<b>High background</b>	Contamination of reagents or samples	Use a clean container before addition into wells.
	Insufficient plates washing	Ensure proper washing of each well
	Too much concentrated detection antibody and Streptavidin-HRP	Ensure proper dilution of detection antibody or conjugate and incubation time.
	Substrate solution or stop solution is not fresh	Use fresh substrate and stop solution.
	Plate left too long before reading on the plate reader	Read on the plate reader right after the experiment.
	Incubation temperature is too high	Decrease incubation temperature of substrate.

<b>Poor standard curve</b>	Samples contain no or below detectable levels of analyte or samples contain analyte concentrations greater than the highest standard point.	If samples are below detectable levels, higher sample volume. If samples are higher than detectable levels, it may require dilution and reanalysis.
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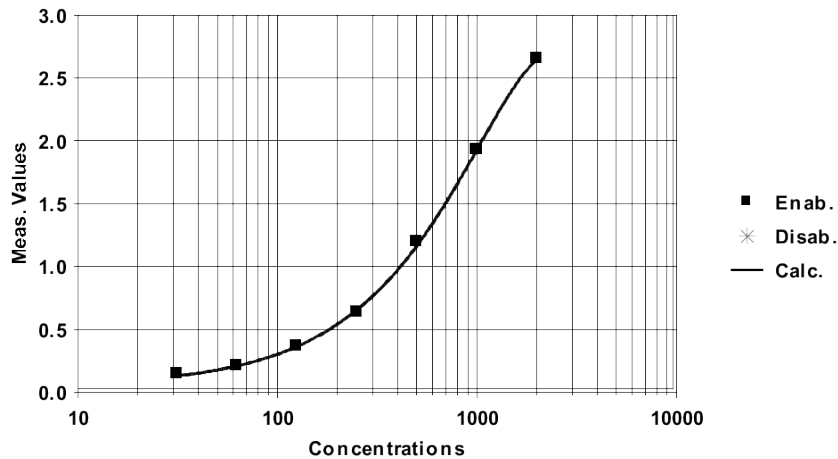
## PLATE LAYOUT

	1	2	3	4	5	6	7	8	9	10	11	12
A												
B												
C												
D												
E												
F												
G												
H												

# CERTIFICATE OF ANALYSIS

**Product** Human TGF-beta 2 ELISA kit, pink-ONE  
**Catalog No.** K0332120P  
**Lot No.** 33232  
**Quantity** 96 tests  
**Storage** 4°C  
**Standard Range** 31.25-2000 pg/ml

ENABLED AND DISABLED MEAN VALUE



**Human TGF-beta 2 (pg/ml)**  
 (3 minutes color development)

## Layout map for calibrators sheet

	Concentration		OD (450nm)
A	Cal_1	2000 pg/ml	2.654
B	Cal_2	1000 pg/ml	1.933
C	Cal_3	500 pg/ml	1.197

D	Cal_4	250 pg/ml	0.637
E	Cal_5	125 pg/ml	0.365
F	Cal_6	62.5 pg/ml	0.213
G	Cal_7	31.25 pg/ml	0.148
H	Black	0 pg/ml	0.064